

Metadata and Data Standards for NIDDK Research Data - The ATLAS-D2K Experience

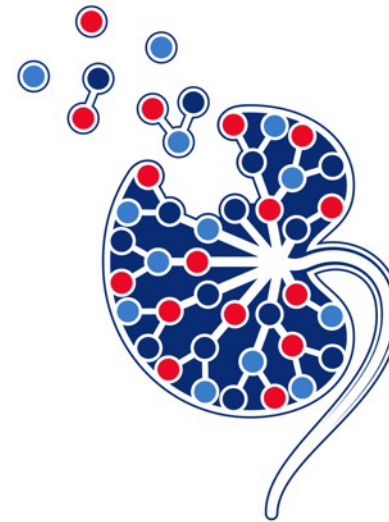
M. Todd Valerius, Ph.D.

Brigham and Women's Hospital / Harvard Medical School

The ATLAS-D2K Center

A kidney and lower urinary tract-focused data discovery hub with access to visualizations and analysis tools.

Bringing GUDMAP & RBK data under one ATLAS that embraces open science and provides links to related consortiums.

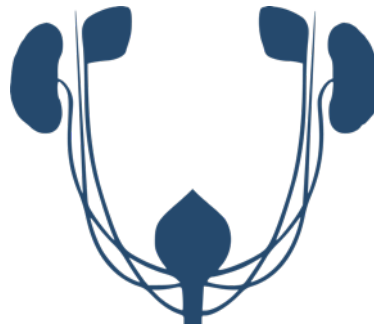


RBK
ReBuilding a Kidney

Overall Aims:



Our long-term goal is to bring complex data into an accessible form for our research community.



Establish connections between molecular data of kidney and lower urinary tract present in GUDMAP, RBK, KPMP, HuBMAP, and the HCA.



Enable researchers of various levels of experience by providing tools to interact with the data.

GenitoUrinary Development Molecular Anatomy Project (GUDMAP) & (Re)Building a Kidney (RBK): Overview

Overarching Program Goal:

- **GUDMAP:** high resolution molecular anatomy of the developing and mature genitourinary system (mouse, human, rat, dog)
- **RBK:** optimize differentiation of human kidney cell types in defined structures, and determine methods to promote kidney repair, to generate or repair nephrons that can function within the kidney (human, human iPSCs, zebrafish)

Number of investigators involved: GUDMAP: 9 RBK: 25

Technology Focus: array of gene expression techniques on tissues, differentiation of stem cells, a range of imaging techniques

Current Gaps? robust anatomical ontologies broadly implemented, metadata standardization, interactive tools for data analysis/data annotation (e.g., cluster data)

ATLAS-D2K Research goals



Example queries:

- a) scRNA-Seq data analysis (rookie/veteran)
- b) GWAS gene list mapped to expression.



Graphical Tools for Genitourinary Data



Integrating molecular and imaging data



Establishing reference datasets



Data harmonization across consortiums



Bioinformatic pipelines and visualization tools

Ontologies and controlled vocabularies

- Why is this important?
 - Gene expression and function occurs in tissues. Consistent use of names removes confusion amongst researchers and **enables** computation of complex queries.
 - Quickly apparent when trying to connect data
 - GUDMAP had generated thousands of wholemount & section *in situ* hybridizations, scored for expression, from two groups. -> **anatomical ontology** needed to connect.

Abler, L.L. *et al.* (2011) *Developmental dynamics : an official publication of the American Association of Anatomists*. <https://doi.org/10.1002/dvdy.22730>.

Georgas, K.M. *et al.* (2015) *Development (Cambridge, England)*. <https://doi.org/10.1242/dev.117903>.

Harding, S.D. *et al.* (2011) *Development (Cambridge, England)*. <https://doi.org/10.1242/dev.063594>.

Henry, G.H. *et al.* (2018) *Cell Rep.* <https://doi.org/10.1016/j.celrep.2018.11.086>.

Little, M.H. *et al.* (2007) *Gene expression patterns : GEP.* <https://doi.org/10.1016/j.modgep.2007.03.002>.

Boolean Search on Scored Expression

Boolean Search

Mus musculus Anatomy Tree

TS17: 10.5 dpc (range 10-11.25 dpc) ▾

metanephric mesenchyme ✕ 🔍 [Expand All](#) [Collapse All](#)

- amniotic cavity (EMAPA:16079)
- C1 dorsal root ganglion (EMAPA:25144)
- cardiovascular system (EMAPA:16104)
- dorsal root ganglion (EMAPA:16668)
- ectoderm (EMAPA:35985)
- endocrine system (EMAPA:35306)
- exocoelomic cavity (EMAPA:16081)
- forelimb bud (EMAPA:16406)
- gland (EMAPA:18425)
- head (EMAPA:31858)
- heart great vessel (EMAPA:36460)
- hindlimb bud (EMAPA:16779)
- intraembryonic coelom (EMAPA:16088)
- left lung rudiment (EMAPA:16729)
- left umbilical vein (EMAPA:36019)
- left vitelline vein (EMAPA:36022)
- limb (EMAPA:16405)
- mesenchyme (EMAPA:16097)
- mesoderm (EMAPA:35987)
- mesothelium (EMAPA:32856)
- mouse (EMAPA:25765)
- musculature (EMAPA:35577)
- musculoskeletal system (EMAPA:32714)
- nervous system (EMAPA:16469)
- perioptic vascular plexus (EMAPA:36465)
- peritoneum parietal mesothelium (EMAPA:16591)
- peritoneum visceral mesothelium (EMAPA:16592)
- pulmonary artery (EMAPA:17008)

p[in "mesonephros" TS16..TS17] AND nd[in "gonad primordium" TS16..TS28] AND f Search Specimen [✕](#) [📄](#) [📥](#) [📧](#) [+](#)

	* Strength	* In Anatomical Source	* Stages		With Pattern	At Location	Actions
	present ▾	mesonephros	From: TS16 ▾	To: TS17 ▾	▾	▾	✕ 🗑️
AND	not detected ▾	gonad primordium	From: TS16 ▾	To: TS28 ▾	▾	▾	✕ 🗑️
AND	present ▾	metanephric mesenchyme	From: TS16 ▾	To: TS28 ▾	▾	▾	✕ 🗑️

Specimen

▼ Gene

Search 🔍

All Records With Value

No Value

Hoxa10

Nampt

Paps2

Tmem100

[Show Details](#)

► Protein

► Tissue (Anatomical Source)

► Expression Region

► Expression Strength

► Species

► Stage

► Chronological Age







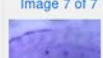





► Assay Type

► Preparation

Search 🔍 25 Items per page ▾

[Clear All](#) [✕ Custom Facets: p\[in "mesonephros" TS1...](#)

Displaying 4 of 4 Records

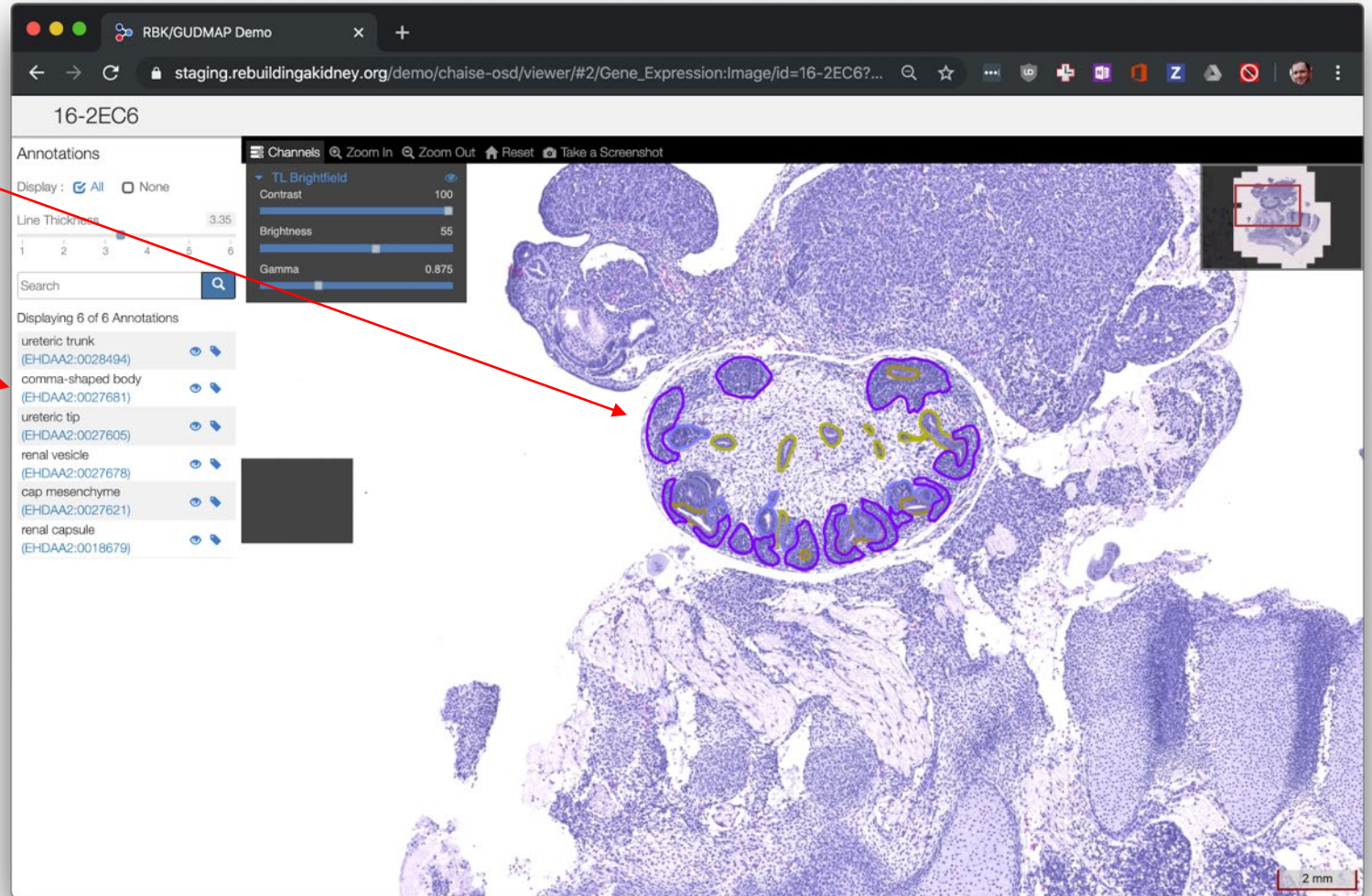
View	RID ↑↓	Imaging Data	Genes ↑↓	Species ↑↓	Stage ↑↓	Anatomical Sources	Assay Type ↑↓	Preparation ↑↓	Principal Investigator ↑↓	Consortium ↑↓	Last Modified Time ↑↓
👁️	N-GJPO	Image 1 of 7 Image 2 of 7 Image 3 of 7    Image 4 of 7 Image 5 of 7 Image 6 of 7    Image 7 of 7 	Tmem100	Mus musculus	TS17	genitourinary system (TS13-TS28)	ISH	wholmount	Melissa H. Little, MCRI	GUDMAP	2019-01-24 20:38:07
👁️	N-GJPE	Image 1 of 6 Image 2 of 6 Image 3 of 6    Image 4 of 6 Image 5 of 6 Image 6 of 6   	Nampt	Mus musculus	TS17	genitourinary system (TS13-TS28)	ISH	wholmount	Melissa H. Little, MCRI	GUDMAP	2019-01-24 20:38:07

Search Results

Anatomical annotation using established terms

Manual annotation of structures

Links to other data through anatomy



Ontologies and controlled vocabularies

- Gene expression and function occurs in tissues. Consistent use of names removes confusion amongst researchers and **enables** computation of complex queries.
- Quickly apparent when trying to connect data
 - GUDMAP had generated thousands of wholemount & section *in situ* hybridizations, scored for expression, from two groups. -> **anatomical ontology** needed to connect.
- To accommodate cross-species data, we use Uberon and Cell Ontology
 - Uberon multi-species anatomy ontology
 - The Cell Ontology
- Healthy adult human tissue focus in **HuBMAP ASCT+B Tables**
 - Data-driven effort lead by Sanjay and enhanced by many.



Recommendation:

Select a source of anatomical and cell type terms that fit your research, use them as a standard, and capture the source of those terms from established ontologies.

Data formats:

Is anything as future proof as plain text?

RAW sequence data is, but there are privacy issues.

- Detailed *sequencing metadata* is produced and captured by computational tools.
- *Biosample metadata* needs to be captured well at the time of experiment.
- Protocols should be well referenced. (consortiums rely on self hosting OR commercial repositories like Protocols.io)

Image data is a mature, poor example

- Center on "open" established standards like OME-TIFF and related formats (e.g., Zarr and OME-NGFF) that capture microscopy/imaging metadata.
 - The benefit: these formats work well with open-source software like ImageJ and QuPath.
 - Images adjusted for publication and presentation are not useful for downstream reuse and quantitative analysis.
- The "biosample metadata" associated with an image needs to be captured EARLY in the process.

Data Curation/Interaction UI: Gene

Direct to expression data associated with this gene

Search for presence of different types of expression data, scored exp. region, etc.

The screenshot shows the RBK/GUDMAP Gene page. The main content is a table of gene results with columns for Gene ID, Gene Symbol, Species, and Description. The table lists genes such as ACTA1, ACTA2, APLNR, AQP1, ASS1, and BMP4. To the right of the table, there are sections for 'Available Expression Data' and 'Representative Images'. The 'Available Expression Data' section lists data types like 'Imaging data', 'Scored expression data', and 'scRNASeq visualization data'. The 'Representative Images' section shows a grid of images for each gene, with labels for different developmental stages (e.g., Fetal 11wk, Fetal 13wk, Fetal 15wk, Fetal 16wk, Fetal 18wk, CS20, CS22, CS23).

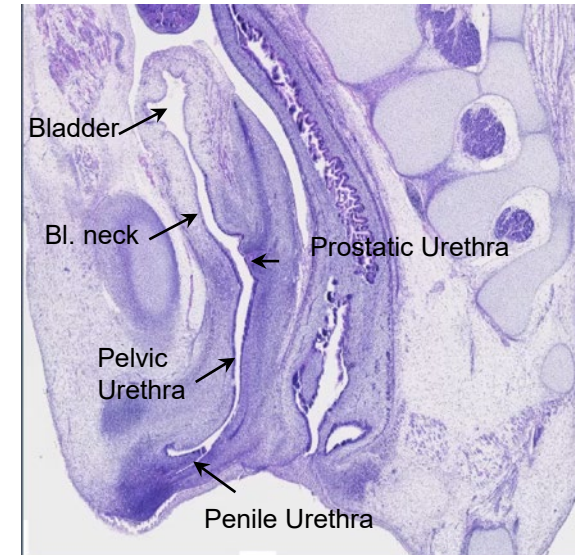
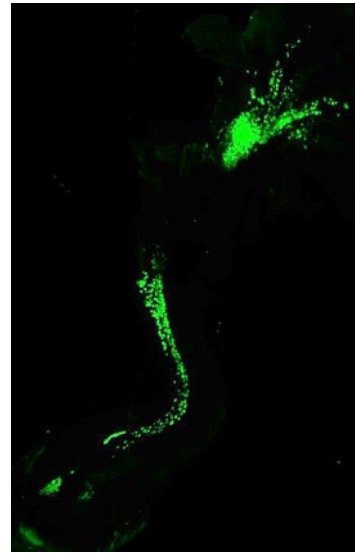
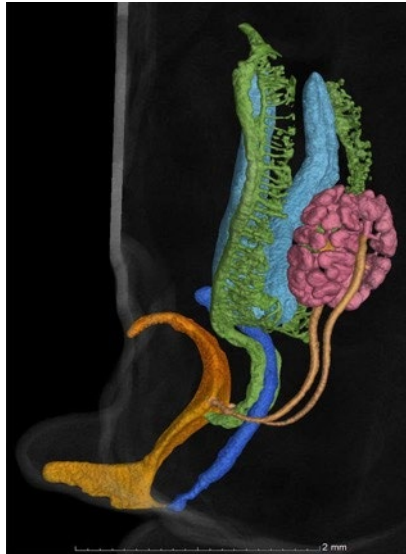
View	Gene ID	Gene Symbol	Species	Description	Available Expression Data	Representative Images
	58	ACTA1	Homo sapiens	actin, alpha 1, skeletal muscle	• Imaging data • Scored expression data • scRNASeq visualization data	Fetal 18wk Fetal 18wk Fetal 13wk Fetal 14wk
	59	ACTA2	Homo sapiens	actin, alpha 2, smooth muscle, aorta	• Imaging data • scRNASeq visualization data	Fetal 11wk Fetal 16wk Fetal 13wk Fetal 15wk
	187	APLNR	Homo sapiens	apelin receptor	• Imaging data • scRNASeq visualization data	CS22 CS23 CS20
	358	AQP1	Homo sapiens	aquaporin 1 (Colton blood group)	• Imaging data • scRNASeq visualization data	Fetal 16wk
	445	ASS1	Homo sapiens	argininosuccinate synthase 1	• Imaging data • scRNASeq visualization data	
	652	BMP4	Homo sapiens	bone morphogenetic protein 4	• Imaging data • scRNASeq visualization data	

Direct to all data associated with this gene

Direct to different specimens associated with this gene

Normal anatomical and structural changes

3-D Mapping of Tissues



Data formats:

Is anything as future proof as plain text?

RAW sequence data is, but there are privacy issues.

- Detailed *sequencing metadata* is produced and captured by computational tools.
- *Biosample metadata* needs to be captured well at the time of experiment.
- Protocols should be well referenced. (consortiums rely on self hosting OR commercial repositories like Protocols.io)

Image data is a mature, poor example

- Center on "open" established standards like OME-TIFF and related formats (e.g., Zarr and OME-NGFF) that capture microscopy/imaging metadata.
 - The benefit: these formats work well with open-source software like ImageJ and QuPath.
 - Images adjusted for publication and presentation are not useful for downstream reuse and quantitative analysis.
- The "biosample metadata" associated with an image needs to be captured EARLY in the process.

Recommendations:

1. Develop a plan to capture biosample metadata and protocol with the sequencing data.
2. Capture and associate biosamples metadata and consolidate on a lossless image file standard.

Data storage and levels of sharing, accessibility to open-source tools.

- **Sequence data – a layered approach**

- "Processed" data is useful to a wider range of researchers.
 - Count files are ready for analysis without computationally intensive genome aligners in HPCs. More researchers can use such data immediately.
- Privacy of participants - what is reasonable to share even with full consent?
 - The ability to de-identify participants from limited sequencing data expands before thoughtful policy will catch up. Think beyond the contractual protection to anticipate while maintaining data availability.
- What intermediate products are available?
 - R objects like Seurat capture analysis decisions for a data generators fine analysis and can be used by less experienced researchers subsequently.
- Processed data more freely shareable, but what happens when references change?
 - As data ages, re-alignment may be necessary as reference genomes change and improve.

Direct linking to data for efficiency with large datasets

- New approach to scientific rigor and reproducibility
 - Data followed from slide to database image
 - "Largest possible" supplementary data
- Collections designed around specific structures

The screenshot shows a web browser window with the URL rebuildingakidney.org. The page title is 'Single-Nucleus versus Single-Cell RNA Sequencing of Adult Mouse Kidney' under the 'COLLECTION' tab. The main content area displays a record with the following fields:

- RID:** 14-4KG6
- Title:** Single-Nucleus versus Single-Cell RNA Sequencing of Adult Mouse Kidney
- Description:** Using adult mouse kidney, we compared single-cell RNA sequencing (scRNA-seq) data generated using the DropSeq platform with single-nucleus RNA sequencing (snRNA-seq) data generated using sNuc-DropSeq, DroNc-seq, and Chromium platforms. We validated snRNA-seq on fibrotic kidney from mice 14 days after unilateral ureteral obstruction (UUO) surgery. This dataset is related to the following publication:
Haojia Wu, Yuhei Kirita, Erinn L. Donnelly and Benjamin D. Humphreys, Advantages of Single-Nucleus over Single-Cell RNA Sequencing of Adult Kidney: Rare Cell Types and Novel Cell States Revealed in Fibrosis, Journal of the American Society of Nephrology, December 2018, ASN.2018090912, DOI: https://doi.org/10.1681/ASN.2018090912
- Require DOI?:** Yes
- Persistent ID:** <https://doi.org/10.25548/14-4KG6>
- Principal Investigator:** Benjamin Humphreys, WUSTL
- Data Provider:** Washington University, St. Louis
- Consortium:** RBK
- Creation Time:** 2018-11-09 14:00:15
- Last Modified Time:** 2018-12-12 17:24:04

Below the record, there is a 'Sequencing Study Collection' section showing 1 result. A table at the bottom provides a summary of the collection:

View	RID ↓↑	Internal ID ↓↑	Title ↓↑	Funding ↓↑	Principal Investigator ↓↑	Data F
	14-4KBA	Mouse_kidney_snRNA_seq	Single-Nucleus versus Single-Cell RNA		Benjamin Humphreys, WUSTL	Washin Univers

Multiple layers with scRNA-seq

1. RAW sequencing files (fastq)
2. Processed gene expression matrix files (txt)
3. R objects of analysis (Rds, e.g., Seurat)
4. Static visualization tools
5. Interactive visualization tools

Direct linking to data for efficiency with large datasets

16-1YZM: Single Cell RNA-Seq data of iPSC-derived Human Kidney Organoids
STUDY

RID 16-1YZM
Internal ID kidney_organoids_scRNAseq
Title Single Cell RNA-Seq data of iPSC-derived Human Kidney Organoids
Summary These files represent single cell RNA-Seq data generated on a 10x Chromium genomics platform from four biological replicates of iPSC-derived human kidney organoids, in two batches, differentiated according to our published protocol (Takasato et al., Nature Protocols 2016). The aggregated human organoid data contains populations representing endothelial cells, podocytes, stroma, nephron, and off-target populations with similarity to neurons.

View	RID ↑	Name ↓	Description ↓	File ↓
	16-3EMT	cello object	Cello object containing TSNE projections for the kidney organoid single cell data.	clist.rds
	16-3EMR	Expression set object	The expression set object for testing VisCello visualisation software.	eset.rds
	16-2D54	Kidney organoids Seurat object	Seurat object containing gene expression information and clustering analysis information.	Organoids_clustered_Seurat.Rds
	16-1ZBR	org4_barcodes		org4_barcodes.tsv
	16-1ZC6	org4_counts		org4_counts.csv.gz
	16-1ZC0	org4_genes		org4_genes.tsv

- New approach to scientific rigor and reproducibility
 - Data followed from slide to database image
 - "Largest possible" supplementary data
 - Collections designed around specific structures
- ## Multiple layers with scRNA-seq
1. RAW sequencing files (fastq)

```
organoid-Little.R  
13 "https://www.gudmap.org/hatrac/resources/rnaseq/study/16-1YZM/study_file  
/2863fb02142e11be217a19aa7004d199:AEVTKSQZG7XDMNUOCGMQOKOMDA?uinit=1"  
14  
15 # Can fetch data from the terminal:  
16  
17 # wget "https://www.gudmap.org/hatrac/resources/rnaseq/study/16-1YZM  
/study_file/2863fb02142e11be217a19aa7004d199:AEVTKSQZG7XDMNUOCGMQOKOMDA  
?uinit=1"  
18  
19 # wget worked well though I then renamed the file (could have done during  
the wget) using mv.  
20  
21 VlnPlot(object = `organoid_Little_Seurat`, do.return = TRUE, point.size  
.use = 0.01, size.x.use = 6, features.plot = c("HNF4A", "GATA3"), nCol =  
1)  
22  
23 VlnPlot(object = `organoid_Little_Seurat`, do.return = TRUE, point.size  
23:1 (Top Level) :  
R Script  
Console Terminal x  
Terminal 1 | mtv9@da02:~/R-projects/organoid-Little  
99% [=====] 1,249  
99% [=====] 1,249  
99% [=====] 1,250  
99% [=====] 1,250  
100% [=====] 1,250  
,565,104 1.43MB/s in 14m 47s  
2019-01-26 13:01:32 (1.34 MB/s) - '2863fb02142e11be217a19aa7004d199:AEVTKSQZG7  
XDMNUOCGMQOKOMDA?uinit=1' saved [1259565104/1259565104]  
[mtv9@da02 organoid-Little]$ ls -l  
total 1493888
```

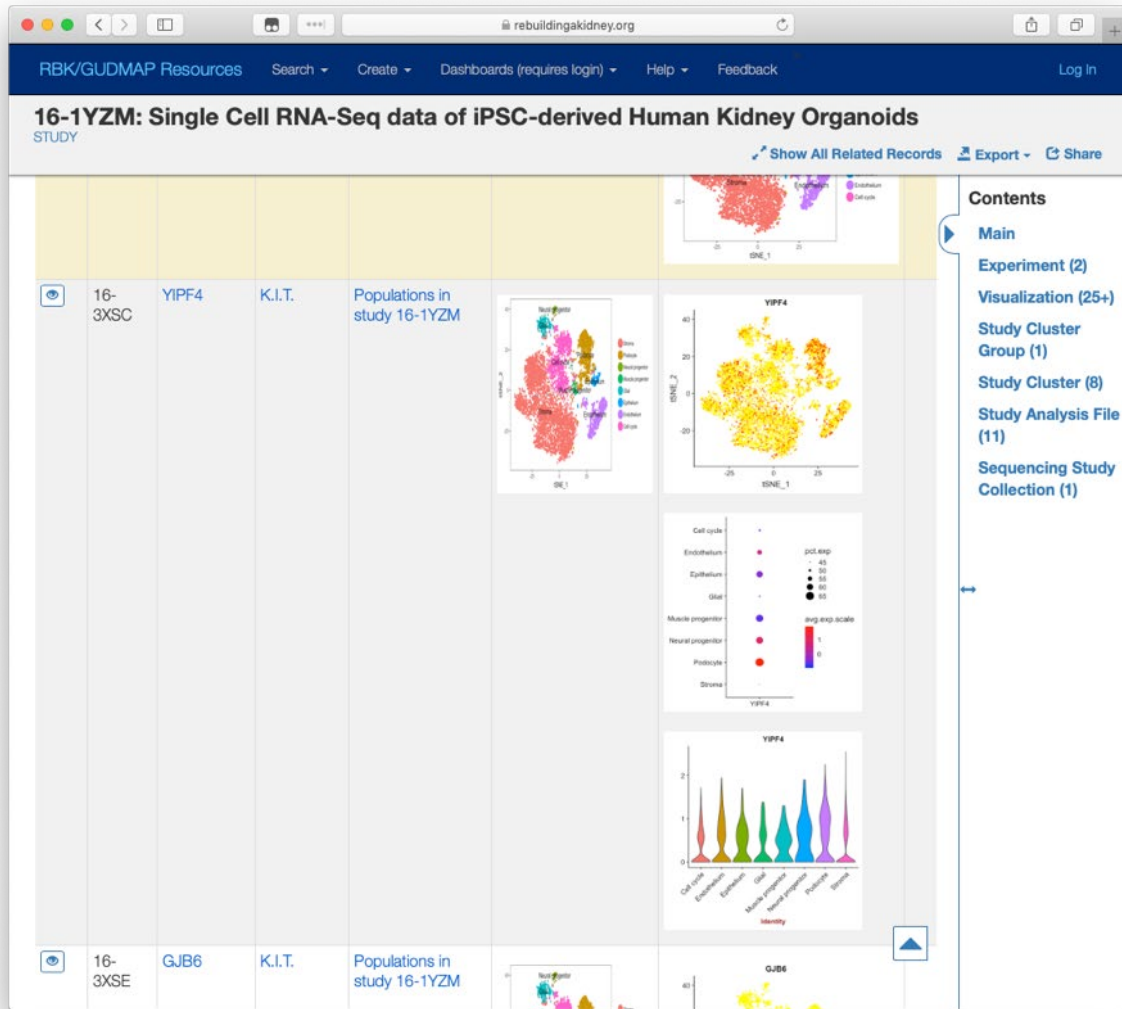
Environment History Connections
Global Environment
Data
colData 7937 obs. of 45 variables
organoid_Little_S. Large seurat (1.7 Gb)

Files Plots Packages Help Viewer
Zoom Export Publish

HNF4A
1.5
1.0
0.5
0.0
0 1 2 3 4 5 6 7 8 9 10 11 12
Identity

GATA3
3
2
1
0
0 1 2 3 4 5 6 7 8 9 10 11 12
Identity

scRNA-seq Visualizations - Accessibility



- New approach to scientific rigor and reproducibility
 - Data followed from slide to database image
 - "Largest possible" supplementary data
- Collections designed around specific structures

Multiple layers with scRNA-seq

1. RAW sequencing files (fastq)
2. Processed gene expression matrix files (txt)
3. R objects of analysis (Rds, e.g. Seurat)
4. Static visualization tools
5. Interactive visualization tools

Data storage and levels of sharing, accessibility to open-source tools.

- **Sequence data – a layered approach**

- "Processed" data is useful to a wider range of researchers.
 - Count files are ready for analysis without computationally intensive genome aligners in HPCs. More researchers can use such data immediately.
- Privacy of participants - what is reasonable to share even with full consent?
 - The ability to de-identify participants from limited sequencing data expands before thoughtful policy will catch up. Think beyond the contractual protection to anticipate while maintaining data availability.
- What intermediate products are available?
 - R objects like Seurat capture analysis decisions for a data generators fine analysis and can be used by less experienced researchers subsequently.
- Processed data more freely shareable, but what happens when references change?
 - As data ages, re-alignment may be necessary as reference genomes change and improve.

mRNA-Seq Reanalysis Progress

# Studies	# Experiments	# Replicates	# Files
49	184	602	851

 **First Round of execution**

Execution Status	Count (# Replicates)	Description
Success	110 (18.3%)	
Error	492 (81.7%)	
- Metadata	311 (51.7%)	<ul style="list-style-type: none"> - Validate: Species, Paired-End, Strandedness, Spikes-in - No metadata or mismatched - Incorrect sequencing type (e.g. ChiP-Seq instead of mRNA-Seq)
- File	181 (30.0%)	<ul style="list-style-type: none"> - Mismatched #Reads of R1 and R2, multiple runs, missing files - Not fastq structure (e.g. fastq+bam)

 **After a few rounds of resolution and execution**

Success	<ul style="list-style-type: none"> - 549 mRNA-Seq replicates (45 studies) - Re-labeled 3 mRNA-Seq replicates to ChiP-Seq (1 study)
Outstanding issues	<ul style="list-style-type: none"> - Missing files: 3 replicates - Conflicting files: 50 replicates (3 studies)

mRNA-Seq QC, Processed Files, and Visualization

Study: Q-Y4GY: Transcriptional profiling of mouse and human nephron progenitors: mouse samples

mRNA QC

View	RID	Replicate	Execution Run	Execution Status	Paired End	Strandedness	Median Read Length	Raw Count	Final Count
	17-CJHW	Q-YSEA (1, 1)	17-CJF6	Success: Run Successful	Paired End	reverse	76	46,264,311	10,593,940
	17-CJIG	Q-YSEE (3, 1)	17-CJFP	Success: Run Successful	Paired End	reverse	76	53,222,569	29,613,404
	17-CJKY	Q-YSEC (2, 1)	17-CJFJ	Success: Run Successful	Paired End	ren			
	17-CME6	Q-YSER (2, 1)	17-CMAP	Success: Run Successful	Paired End	ren			
	17-CMEJ	Q-YSEW (1, 1)	17-CMAJ	Success: Run Successful	Paired End	ren			
	17-CMFA	Q-YSF6 (3, 1)	17-CMAT	Success: Run Successful	Paired End	ren			
	17-CMFP	Q-YSEP (1, 1)	17-CMBB	Success: Run Successful	Paired End	reverse	76	51,000,000	27,000,000
	17-CMGE	Q-YSEY (2, 1)	17-CMBE	Success: Run Successful	Paired End	reverse	76	55,527,687	32,489,938

Replicate-level Submitted File

View	RID	Experiment	Replicate	Species	Caption	File Type	File	Notes	Curation Status
	Q-Y52J	Q-YADP: mMARIS_Six2-	Q-YSF6 (3, 1)	Mus musculus	Mus musculus mMARIS_Six2-replicate 3 alignment	bam	mMARIS_Six2_#3_sorted.bam		Release
	Q-Y52M	Q-YADP: mMARIS_Six2-	Q-YSF6 (3, 1)	Mus musculus	Mus musculus mMARIS_Six2-replicate 3 visualization track	bigWig	mMARIS_Six2_#3_normalized.profile.bigwig		Release
	Q-Y52R	Q-YADP: mMARIS_Six2-	Q-YSF6 (3, 1)	Mus musculus	Mus musculus mMARIS_Six2-replicate 3 R reads	FastQ	mMARIS_Six2_#3_R2.fastq.gz		Release
	Q-Y532	Q-YADP: mMARIS_Six2-	Q-YSF6 (3, 1)	Mus musculus	Mus musculus mMARIS_Six2-replicate 3 F reads	FastQ			Release

Replicate-level Processed Files

View	RID	Replicate	Execution Run	File URL	File Type
	17-CJHY	Q-YSEA (1, 1)	17-CJF6	Q-YSEA_tpmTable.csv	csv
	17-CJJO	Q-YSEA (1, 1)	17-CJF6	Q-YSEA_sorted.deduped.bigwig	bigWig
	17-CJLJ	Q-YSEA (1, 1)	17-CJF6	Q-YSEA_sorted.deduped.bam	bam
	17-CJLJ	Q-YSEA (1, 1)	17-CJF6	Q-YSEA_sorted.deduped.bam.bai	bai
	17-CJKJ	Q-YSEE (3, 1)	17-CJFP	Q-YSEE_sorted.deduped.bam	bam
	17-CJMK	Q-YSEE (3, 1)	17-CJFP	Q-YSEE_sorted.deduped.bigwig	bigWig
	17-CJMP	Q-YSEE (3, 1)	17-CJFP	Q-YSEE_sorted.deduped.bam.bai	bai
	17-CJMR	Q-YSEE (3, 1)	17-CJFP	Q-YSEE_tpmTable.csv	csv
	17-CJMO	Q-YSEC (2, 1)	17-CJFJ	Q-YSEC_sorted.deduped.bigwig	bigWig
	17-CJMJ	Q-YSEC (2, 1)	17-CJFJ	Q-YSEC_sorted.deduped.bam.bai	bai
	17-CJMA	Q-YSEC (2, 1)	17-CJFJ	Q-YSEC_sorted.deduped.bam	bam
	17-CJMA	Q-YSEC (2, 1)	17-CJFJ	Q-YSEC_tpmTable.csv	csv
	17-CJME	Q-YSER (2, 1)	17-CMAP	Q-YSER_tpmTable.csv	csv

QC data (including execution status)

User submitted seq files and hub-processed analysis files are all accessible

Study: Q-Y4GY: Transcriptional profiling of mouse and human nephron progenitors: mouse samples

TPM Expression

Gene: Group By: Scale:

Interact visualization of TPM expression (group by Experiment, Anatomical Source, Stage, etc)

<https://dev.gudmap.org/id/Q-Y4GY> (dev server only)

Execution Run: 17-CMBM

Workflow: BICF mRNA Replicate v2.0.0
 Reference Genome: GRCh38.p6-M25
 Input Bag: Q-YSEJ_inputBag_20210413.zip
 Output Bag: Q-YSEJ_outputBag_20210413.zip
 Execution Status: Success
 Execution Status Detail: Run Successful
 Last Modified Time: 2021-04-14 04:31:47

mRNA QC

View	RID	Replicate	Execution Status	Paired End	Strandedness	Median Read Length	Raw Count	Final Count	Notes
	17-CMGT	Q-YSEJ (2, 1)	Success: Run Successful	Paired End	reverse	76	59,300,385	33,270,091	

Output Bag

View	RID	File URL	Bag Type	Notes	File Bytes	File MD5	File Creation Time
	17-CMH4	Q-YSEJ_Output_Bag.zip	mRNA_Replicate_Analysis		479,984	c2af9f72a439a0416930219db2bf7da	2021-04-14 06:31:44

Individual execution run contains workflow definition, version, source code URL, input and output for reproducibility

<https://www.gudmap.org/id/17-BMCM>

<https://www.gudmap.org/id/Q-Y4GY>

Data storage and levels of sharing, accessibility to open-source tools.

• **Sequence data – a layered approach**

- "Processed" data is useful to a wider range of researchers.
 - Count files are ready for analysis without computationally intensive genome aligners in HPCs. More researchers can use such data immediately.
- Privacy of participants - what is reasonable to share even with full consent?
 - The ability to de-identify participants from limited sequencing data expands before thoughtful policy will catch up. Think beyond the contractual protection to anticipate while maintaining data availability.
- What intermediate products are available?
 - R objects like Seurat capture analysis decisions for a data generators fine analysis and can be used by less experienced researchers subsequently.
- Processed data more freely shareable, but what happens when references change?
 - As data ages, re-alignment may be necessary as reference genomes change and improve.

Recommendations:

1. Consider asking your core to use programmatically published base processing pipelines.
2. Publish these AND analysis code to a coding repository. (Contemporaneous documentation saves time later.)
3. Consider what intermediate layers should be protected (RAW versus counts).
4. Publish analysis intermediates.

ATLAS-D2K Team

